

Compositions comprising polypeptides

The present invention relates to compositions comprising polypeptides, especially polypeptides capable of specifically binding to predetermined antigens via epitopes on said antigens. A preferred composition is a pharmaceutical composition. The present invention also relates to a method of producing an enriched composition in which the amount of a polypeptide in monomeric form has been enriched relative to other multimeric forms of the polypeptide. The present invention also relates to an enriched composition produced by the above method. The present invention further relates to methods for the prevention, treatment or amelioration of various diseases. Finally, the present invention relates to the use of compositions for producing a medicament for the prevention, treatment or amelioration of these various diseases.

With the advent of standardized methods of producing recombinant polypeptides and proteins, such recombinant species are increasingly being employed as the active therapeutic agents in pharmaceutical compositions for the treatment of human disease states. Given the number of companies, research organizations and university laboratories engaging in the development of recombinant therapeutic polypeptides and proteins, the number of medicinal compositions in which the therapeutic effect is attributable to a recombinantly produced polypeptide or protein can only be expected to increase in the future.

Due to their high binding selectivity and affinity, the immunoglobulins ("Igs"), or antibodies, represent one especially relevant class of proteins of high therapeutic potential. Of particular interest in recent years have been recombinantly produced single chain antibodies in both monospecific and bispecific forms. Monospecific single chain antibodies are disclosed for example in US 4,946,778. A bispecific single chain antibody is disclosed for example in US 5,091,513. Such bispecific single chain antibodies can be of particular therapeutic relevance, since the two distinct functionalities within such a species can efficiently and selectively bring two distinct epitopes, that is in most cases two distinct antigens *in vivo* together spatially. Due to the fact that a bispecific single chain molecule unites two antigen binding sites on a single contiguous polypeptide chain, such molecules overcome the problems of recombinant producibility experienced for full Igs due for example to the latter's comprising an Fc portion.

Of particular therapeutic interest has been the development of recombinantly produced antibodies, for example bispecific single chain antibodies, which are capable of specifically binding to the human CD3 antigen.

5 The human CD3 antigen is present on both helper T cells and cytotoxic T cells. The latter, namely cytotoxic T cells, are responsible for the killing of invading or infected cells against which the cytotoxic T cells have been activated. Human CD3 denotes an antigen which is expressed on T cells as part of the multimolecular T cell complex and which comprises three different chains: CD3-epsilon, CD3-delta and CD3-gamma.

10 The activation of the cytotoxic potential of T cells is a complex phenomenon which requires the interplay of multiple proteins. The T cell receptor ("TCR") protein is a membrane bound disulfide-linked heterodimer consisting of two different glycoprotein subunits. The TCR recognizes and binds foreign peptidic antigen which itself has been
15 bound by a member of the highly diverse class of major histocompatibility ("MHC") proteins and has been presented, bound to the MHC, on the surface of antigen presenting cells ("APCs").

Although the variable TCR binds foreign antigen as outlined above, signalling to the T
20 cell that this binding has taken place depends on the presence of other, invariant, signalling proteins associated with the TCR. These signalling proteins in associated form are collectively referred to as the CD3 complex.

In summary, the activation of T cell cytotoxicity normally depends first on the binding of
25 the TCR with an MHC protein, itself bound to foreign antigen, located on a separate cell. Only when this initial TCR-MHC binding has taken place can the CD3-dependent signalling cascade responsible for T cell clonal expansion and, ultimately, T cell cytotoxicity ensue.

30 However, it has previously been found that certain recombinantly produced, polypeptidic antigen binding sites which specifically bind to at least part of the human CD3 antigen have the ability to activate T cells to exert a cytotoxic effect on other cells in the absence of independent TCR-MHC binding. This means that T cells may become cytotoxically active in a clonally independent fashion, i.e. in a manner which is
35 independent of the specific TCR clone carried by the T cell. This allows an activation of

the entire T cell compartment rather than only specific T cells of a certain clonal identity. Such molecules have been disclosed in WO 99/54440; Mack, J. Immunol. (1997) 158, 3965-70; Mack, PNAS (1995) 92, 7021-5; Kufer, Cancer Immunol. Immunother. (1997) 45, 193-7; Löffler, Blood (2000) 95, 2098-103; Brühl, J. Immunol. (2001) 166, 2420-6.

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The type of biological activity described above, i.e. the ability of a polypeptide to selectively (re)direct the cytotoxic potential of cytotoxic T cells against predetermined target cells such that the latter become lysed, can be of great therapeutic relevance. Specifically, compositions of such polypeptides as those described in the previous

10 paragraph can be and have been effectively used as part of a regimen of therapy entailing the destruction of target cells associated with particular diseases. In particular, such diseases include cancerous states in which transformed cells are the target cells destined for destruction.

15 In addition to having the sort of biological activity described above, i.e. the ability to direct the cytotoxicity of T cells to target cells intended for destruction, compositions comprising polypeptides of the sort described above will often manifest other additional types of biological activities unrelated to the lysis of target cells. Such additional biological activities may or may not be beneficial and, if such a composition is intended

20 for administration to a patient, stand to complicate the constructing of a therapeutic regimen. It would therefore be desirable to eliminate such additional types of biological activities to the greatest extent possible in such a composition, so that the type of biological activity manifested by the resulting composition remains as homogeneous as possible.

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It is therefore an object of the invention to provide a composition which overcomes the above difficulties.

Accordingly, the present invention provides a composition with a polypeptide. The

30 polypeptide comprises at least two antigen binding sites, wherein said at least two antigen binding sites are located on a single polypeptide chain, and wherein

- one of said at least two antigen binding sites specifically binds the human CD3 antigen;
 - said polypeptide may exist in both monomeric form and multimeric form, said
- 35 monomeric form being said single polypeptide chain (with the at least two antigen

binding sites) and said multimeric form comprising at least two of said single polypeptide chains non-covalently associated with one another, thereby comprising at least four antigen binding sites; and

- said multimeric form of said polypeptide constitutes no more than 5% of the total weight of the combined monomeric and multimeric forms of said polypeptide.

The terms "multimeric polypeptide", "polypeptide in multimeric form", "multimer", etc as used herein are equivalent terms and are contemplated as meaning (i) different isoforms within a population of polypeptide molecules multimeric to the same degree (e.g. different dimeric isoforms), and/or (ii) a population of polypeptide molecules which are multimeric to different degrees (e.g. dimers, trimers, etc.).

The term "antigen binding site" is to be understood as a portion of secondary and/or tertiary polypeptide structure which specifically binds an antigen of interest in a non-covalent manner via an epitope of the antigen. Hereinafter, it should be borne in mind that antigens are bound via a specific epitope or via specific epitopes of such antigens. "Specific" binding denotes the ability to discriminate between different antigens as potential binding partners to such an extent that, from a pool of a plurality of different antigens as potential binding partners, only the antigen of interest is bound, or is significantly bound. Within the meaning of the invention, an antigen is "significantly" bound when, from among a pool of equally accessible different antigens as potential binding partners, the antigen of interest is bound at least 10-fold, preferably 50-fold, most preferably 100-fold or greater more frequently (in a kinetic sense) than other antigens which are not the antigen of interest.

Whereas one of the at least two antigen binding sites of the polypeptide comprised in the composition of the invention specifically binds the human CD3 antigen, the at least one other antigen binding site of this polypeptide is allowed to specifically bind any other antigen(or epitope) of interest ("target antigen"). Preferably, the target antigen is an antigen expressed on the surface of a cell, wherein the cell expressing the target antigen/epitope may be a free cell, such as a lymphocyte in the bloodstream, or may form part of a solid tissue. In this manner, the polypeptide comprised in the composition of the invention may with one arm (i.e. one antigen binding site, or the "target antigen binding site") specifically bind to the target antigen, while a/the second arm (i.e. another/the other antigen binding site, or the "effector antigen binding site") of the

polypeptide comprised in the composition specifically binds to and activates, via the human CD3 antigen, a cytotoxic T cell in a clonally independent fashion as described above. In this manner, the polypeptide comprised in the composition according to the invention may be generally employed as part of a therapeutic regimen to specifically
5 destroy, via the cytotoxic T cell, a certain cell type.

As implied above, the polypeptide comprised in the composition according to the invention is thus biologically active. The terms "biologically active" and "biological activity" as used herein denote the nature of an effect caused by the polypeptide
10 comprised in the composition according to the invention when said polypeptide is placed in an *in vitro*, *ex vivo* or *in vivo* setting. As used herein, biological activity therefore refers to types of biological effects elicited rather than a certain effect's magnitude.

It has been surprisingly found that the biological activity of the monomeric form of the polypeptide comprised in the inventive composition is much more homogeneous than that of the multimeric form of this polypeptide. That is to say the monomeric form of the polypeptide demonstrates a single type of biological activity (i.e. activation and redirection of the cytotoxic activity of T cells against target cells intended for
20 destruction), whereas the multimeric form, for example the dimeric form of the polypeptide demonstrates multiple types of biological activity which are different than that manifested by the monomeric form of the polypeptide.

Without being bound by theory, it is believed that the greater diversity of biological
25 activity observed for the multimeric form of the polypeptide comprised in the inventive composition might be due at least in part to the greater number of modes for molecular association available for the multimer as compared to the monomer. That is to say that statistically, there exist a greater number of ways a multimeric species composed of a plurality of single polypeptide chains may associate and become folded than exist for
30 the corresponding monomeric species composed of only one single polypeptide chain. This idea is borne out by a number of findings of the inventors and are discussed in detail hereinafter.

The monomeric species of the polypeptide comprised in the inventive composition
35 exhibits a single biological activity. As explained above, this is the ability to recruit the

cytotoxic T cells ("CTLs") against other cells which are not CTLs, and which bear on their surface an antigen which is specifically bound by the/a target antigen binding site.

While also partially manifesting a biological activity as observed for the monomeric species, one or more of the multimeric species of said polypeptide also give rise to additional biological activities. It was for example observed that the multimeric polypeptide species led to a decrease in the number of CTLs present in a sample. While not being bound by theory, the inventors believe that this biological activity is likely due to intermolecular association of at least two molecules of monomer polypeptide via their respective antigen binding sites. In this way, a multimeric species is formed in which, for example, the target antigen binding sites mutually engage one another, and thus become unavailable for binding target antigen, whereas each effector antigen binding site specific for the human CD3 antigen remains free to bind a respective CD3 antigen. In this way, a species is formed which is capable of specifically binding at least two distinct molecules of the human CD3 antigen by identical epitopes. Such a species would be capable of simultaneously binding at least two separate CTLs, a scenario in which one of these at least two CTLs might exert its cytotoxic effect on any other of the at least two CTLs. This type of biological activity, in which other cells than the target cells intended for destruction (i.e. the cytotoxic T cells themselves) are lysed, stands to decrease the overall number of CTLs present in a sample. This stands to decrease the number of such cytotoxic T cells available for participating in the type of biological activity manifested by the monomeric species, namely the selective destruction, via T cell mediated lysis, of diseased target cells.

In addition, the inventors have recognized that the multimeric forms of the polypeptide as comprised in the inventive composition are able to activate CTLs even in the absence of other types of non-CTL cells. Normally, the monomeric species of the polypeptide comprised in the inventive composition activates the cytotoxic potential of CTLs only in the presence of the cells ("target cells") displaying the antigen which is bound by the target antigen binding site, which cells are accordingly intended for destruction by the CTLs. Activation of CTLs by the polypeptide of the instant composition only in the presence of such target cells advantageously prevents a possible misdirection of cytotoxic activity of CTLs to non-target cells not intended for destruction.

The inventors have also found that the tendency to form a multimeric species, especially a dimeric species, is a property of this class of polypeptides in general, namely single polypeptide chains comprising both a binding site for the human CD3 antigen and a binding site for another target antigen other than the human CD3 antigen.

5 The additional biological activities above may therefore be expected for any polypeptide of this sort, regardless of the specificity of the target antigen binding site.

As follows from the above explanations, a composition comprising only a minimal, controlled amount of polypeptide in multimeric form and wherein total polypeptide is
10 substantially in the monomeric form will demonstrate a more homogeneous biological activity than a composition containing a greater amount of multimeric polypeptide. By prescribing an upper limit for the amount of multimeric polypeptide in the composition of the invention, a composition is obtained for which the degree of homogeneity in biological activity is controlled and predictable. Controllability and predictability of
15 biological activity are two features which are preferable for compositions contemplated for administration as part of a therapeutic regimen.

According to one embodiment of the composition according to the invention, the multimeric form of the polypeptide constitutes no more than 4%, preferably no more
20 than 3%, more preferably no more than 2%, even more preferably no more than 1%, yet more preferably no more than 0.5% of the total combined weight of polypeptide in both monomeric and multimeric forms in the composition. Most preferably, the multimeric forms of the polypeptide constitute only or even less than the detectable limit of the multimeric forms of the polypeptide in the composition, the vast majority of polypeptide
25 being present in the composition in a monomeric form.

The terms "detectable limit" and "detection limit" as used herein are equivalent terms and are to be understood as denoting an amount of multimeric polypeptide in the instant composition below which no detection of said multimeric polypeptide at all is possible,
30 even when applying the most stringent assay with its most stringent conditions. Suitable methods for determining the amount of multimeric polypeptide in the instant composition include any method of detecting polypeptide species, for example by non-denaturing polyacrylamide gel electrophoresis wherein proteins are stained in the gel with Coomassie brilliant blue or silver nitrate, by Western blot analysis or chromatographic
35 methods such as size exclusion HPLC. Preferably, monitoring the amount of multimeric

polypeptide present in the composition can best be accomplished by analytical size exclusion HPLC. By the nature of the term, the "detectable limit" will depend on the sensitivity of the particular detection method used to assay the amount of multimeric form of polypeptide present in a given composition. In addition, the "detectable" limit will
5 understandably depend on how stringently the assay parameters are applied for a given method of choice.

In a further embodiment, the multimeric form of the polypeptide as described above is exclusively the dimeric form of the polypeptide. The "dimeric form" is to be understood
10 as a species comprising two single polypeptide chains, wherein the two single polypeptide chains are non-covalently associated with one another.

Contemplated is a composition comprising a polypeptide which itself comprises two antigen binding sites, and wherein each antigen binding site comprises a variable region
15 from a heavy chain of an antibody (VH) and a variable region from a light chain of an antibody (VL), each VH/VL pair having specificity for a different epitope, preferably for a different antigen, one of which is the human CD3 antigen. The VH and VL regions within a given antigen binding site may be derived from the same or different antibodies. The anti-CD3 binding site may be located at either the N- or C-terminus of the polypeptide.
20 Within the meaning of the instant invention, "VH/VL" or "VH/VL pair" is to be understood as denoting any order of connectivity; either VH-VL or VL-VH. While direct covalent (peptide) attachment of the C-terminal amino acid of a VH or VL region to the N-terminal amino acid of a VL or VH region, respectively, is theoretically possible, one of ordinary skill in the art will understand that such a direct peptide linkage often confers too few
25 spatial degrees of freedom to allow the VH and VL region to associate such that their respective CDR regions can form a single unified antigen binding site. One of skill in the art will therefore understand that such non-covalent association of VH and VL regions consistent with the maintenance of the ability to specifically bind an antigen of choice will often make the inclusion of a peptide linker interposed between the VH and VL
30 regions preferable. Such a peptide linker may take the form of linkers disclosed in the art, for example in EP 0 623 679 B1, US 5,258,498, EP 0 573 551 B1 and US 5,525,491.

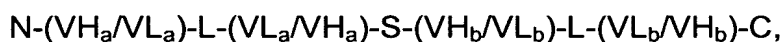
One of skill in the art will appreciate that such a molecule might be expected to form a
35 number of different dimeric forms. It might for instance be expected that the VH and VL

regions making up the target antigen binding site of one monomeric polypeptide molecule would associate in a linear, anti-parallel fashion with the respective VL and VH regions making up the target antigen binding site of another monomeric polypeptide molecule. This would yield a dimeric polypeptide in which the two antigen binding sites specific for the human CD3 antigen would remain free to specifically bind to two separate human CD3 antigens. It might also be expected that the VH and VL regions making up the CD3 antigen binding specificity of one monomeric polypeptide molecule would associate in a linear anti-parallel fashion with the respective VL and VH regions making up the CD3 antigen binding specificity of another monomeric polypeptide molecule. This would yield a dimeric polypeptide in which the two antigen binding sites specific for the target antigen would remain free to specifically bind to two separate target antigens. Also contemplated are pairings between either VH and/or VL regions from the target antigen binding site in one monomeric polypeptide molecule with either VH and/or VL regions from the effector antigen binding site specific for the human CD3 antigen in another monomeric polypeptide molecule. Here, one might expect that the resulting dimeric polypeptide molecule would retain the ability to at least partially bind to each of the human CD3 antigen and the target antigen in a specific manner. The above examples are not limiting in terms of the different species of dimeric polypeptide which may be formed by the polypeptide comprised in the composition of the invention. Clearly, a plurality of different dimeric species can be contemplated, possibly explaining the variegated biological activity observed for multimeric, in particular for dimeric polypeptide.

According to another embodiment of the invention, the composition may comprise polypeptides in which a single antigen binding site comprises two non-covalently associated VH regions on the same polypeptide chain, the two VH regions being separated by a peptide linker as described above, or two non-covalently associated VL regions on the same polypeptide chain, the two VL regions being separated by a peptide linker as described above.

It is envisioned that the VH and/or VL regions of a given antigen binding site may be derived from different sources, for example from two different monoclonal antibodies which may or may not originate from two organisms of the same species, or may be modified (i.e. chimeric, truncated, humanized, deimmunized, etc.).

In an especially preferred embodiment, the polypeptide comprised in the instant composition comprises two antigen binding sites, wherein each antigen binding site comprises one VH and one VL region. In this embodiment, the two antigen binding sites are covalently connected to one another through a short peptide spacer, and each antigen binding site specifically binds a different antigen. As such, a polypeptide according to this embodiment would be represented by the generic formula



where:

- a respective pair “VH/VL” or “VL/VH” represents a mutually exclusive option for choosing either VH or VL at that position;
- “a” and “b” (in subscript) represent specificity for antigen a and b, respectively;
- “L” represents a peptide linker covalently connecting a respective VH and VL or VL and VH within a given single antigen binding site, as discussed above;
- “S” represents a peptide spacer, which is a polypeptide region covalently connecting the antigen binding site specifically binding antigen a with the antigen binding site specifically binding antigen b; and
- “N” and “C” represent the respective N- and C-termini of the polypeptide.

As such, the present embodiment envisions a composition as set out herein comprising a polypeptide with two distinct antigen binding sites, wherein each antigen binding site comprises a VH region and a VL region connected by a peptide linker, and wherein the two antigen binding sites are connected through a single polypeptide spacer. Thus a single polypeptide chain is created on which two antigen binding sites of different specificities are located. One of skill in the art will recognize a species of this general form as a “bispecific single chain antibody”.

It is within the scope of the composition of the invention that the polypeptide comprised therein and as represented by the generic formula above may optionally include other functionalities such as a His-tag or a Flag-tag or other forms of functional labels.

In one particularly preferred embodiment of the invention the composition comprises a polypeptide in which the other of the at least two antigen binding sites, i.e. the target antigen binding site, specifically binds the human CD19 antigen. The human CD19

antigen is expressed in the whole human B lineage from the pro B cell to the mature B cell, it is not shed, is uniformly expressed on all lymphoma cells, and is absent from stem cells. Thus, a composition according to this embodiment, namely one comprising a polypeptide with an antigen binding site which specifically binds the human CD3 antigen as well as an antigen binding site which specifically binds the human CD19 antigen, is of great potential value as a therapeutic. The biological activity of the monomeric form of the polypeptide comprised in a composition advantageously recruits the cytotoxic potential of T cells against B cells in a subject (as explained above). By controlling the multimer:monomer ratio of polypeptide as set out above, a composition is obtained which can advantageously be used to treat B cell-related disorders in an extremely controlled and therefore therapeutically efficacious manner.

Especially preferred is a composition in which the polypeptide with binding specificities for both the human CD3 antigen and the human CD19 antigen has an amino acid sequence equivalent to, or substantially equivalent to any one of those set out in SEQ ID NOs: 1-6 as follows:

- Schematic representation of SEQ ID NO 1: VL(CD19)-L-VH(CD19)-S-VH(CD3)-L-VL(CD3);
- Schematic representation of SEQ ID NO 2: VH(CD19)-L-VL(CD19)-S-VH(CD3)-L-VL(CD3);
- Schematic representation of SEQ ID NO 3: VH(CD3)-L-VL(CD3)-S-VH(CD19)-L-VL(CD19); or
- Schematic representation of SEQ ID NO 4: VH(CD3)-L-VL(CD3)-S-VL(CD19)-L-VH(CD19),
- Schematic representation of SEQ ID NO 5: VL(CD3)-L-VH(CD3)-S-VH(CD19)-L-VL(CD19),
- Schematic representation of SEQ ID NO 6: VL(CD3)-L-VH(CD3)-S-VL(CD19)-L-VH(CD19),

wherein:

- VH(CD19) and VL(CD19) represent a VH region and a VL region, respectively, which associate with one another to form an antigen binding site specifically binding the CD19 antigen via an epitope of the CD19 antigen;
- VH(CD3) and VL(CD3) represent a VH region and a VL region, respectively, which associate with one another to form an antigen binding site specifically binding the CD3 antigen via an epitope of the human CD3 antigen;

- “L” and “S” are as defined above.

Within this embodiment, the term “substantially equivalent to” is understood to comprise amino acid sequences homologous to any of SEQ ID NOs: 1-6 by at least 70%, based on a comparison of primary amino acid sequence. Such degrees of homology may be determined by standard sequence alignment programs such as Vector NTI (InforMax™, Maryland, USA). Such programs compare aligned sequences on an amino acid-by-amino acid basis, and can be set to various levels of stringency for the comparison (e.g. identical amino acid, conservative amino acid substitution, etc.). Within the meaning of this embodiment, two amino acids in question are considered as being “homologous” when they are either identical to one another or conservative substitutions of one another. By way of non-limiting example, two different amino acids belonging to the class of lipophilic amino acids would be considered homologous in the sense of this embodiment, even if these two amino acids were not identical, whereas a lipophilic amino acid on the one hand and a charged acidic amino acid on the other hand would not be considered homologous.

In another preferred embodiment of the invention the composition comprises a polypeptide in which the other of the at least two antigen binding sites, i.e. the antigen binding sites which does not specifically bind the human CD3 antigen, specifically binds the human EpCAM (“Epithelial cell adhesion molecule”, also called 17-1A antigen, KSA, EGP40, GA733-2, ks1-4 or esa) antigen. EpCAM is a 40 kDa membrane integrated glycoprotein of 314 amino acids with specific expression in certain epithelia and on many human carcinomas. EpCAM has been shown in various studies to be beneficial in diagnosis and therapy of various carcinomas. Furthermore, in many cases, tumor cells were observed to express EpCAM to a much higher degree than their parental epithelium or less aggressive forms of said cancers.

In order to obtain a composition according to the invention starting from a composition comprising polypeptide in both monomeric and multimeric form, it is often necessary to adjust the amount (i.e. weight present in the composition) of polypeptide in monomeric form relative to the amount (i.e. weight present in the composition) of polypeptide in multimeric form. As the weight of the polypeptide in multimeric form in untreated compositions, for example cell harvest lysates obtained following protein expression, will often exceed 5% of the total weight of the combined monomeric and multimeric

forms of the polypeptide, it will often be necessary to enrich the content of the polypeptide in monomeric form relative to the content of the polypeptide in multimeric form to obtain the composition of the invention. In general, possibilities include high resolution ion-exchange HPLC, high resolution size exclusion chromatography, gel purification, control of protein expression conditions (e.g. choice of expression host, growth conditions applied to host, expression vector used, type of promoter used, etc.). Advantageous particulars are provided in the examples appended hereto.

In order to accomplish the enrichment mentioned above, another aspect of the invention provides a method of producing a composition in which the amount of a polypeptide in monomeric form has been enriched relative to the amount of said polypeptide in multimeric form. The method comprises the following steps:

a) providing the composition comprising said polypeptide in both multimeric and monomeric form;

b) isolating said polypeptide in both multimeric and monomeric form from said composition, said isolating accomplished by

(b1) applying said composition to a first chromatographic material comprising a metal ion;

(b2) removing any components of said composition which have not bound to said first chromatographic material by washing said first chromatographic material with a first buffer; and

(b3) eluting said polypeptide in both multimeric and monomeric form from said first chromatographic material by applying imidazole to said first chromatographic material in a concentration of at least 60 mM;

(b4) collecting a first eluate comprising said polypeptide in multimeric form and said polypeptide in monomeric form;

c) performing a precursor step that is preparatory for the separation of said polypeptide in multimeric form from said polypeptide in monomeric form to occur in step (d), said precursor step accomplished by

(c1) applying said first eluate to a second chromatographic material, which is an ion exchange material;

(c2) removing any components of the first eluate which have not bound to said second chromatographic material by washing said second chromatographic material with a second buffer;

(c3) eluting said polypeptide in multimeric and monomeric form from said second chromatographic material by applying sodium chloride to said second chromatographic material in a concentration of at least 200 mM;

5 (c4) collecting a second eluate;

d) performing a separation of said polypeptide in multimeric form from said polypeptide in monomeric form, said separation accomplished by

(d1) applying said second eluate to a third chromatographic material allowing separation on the basis of molecular weight;

10 (d2) translocating components of the applied second eluate along said third chromatographic material by applying a running buffer to said third chromatographic material;

(d3) collecting a third eluate in fractions;

e) analyzing said fractions of said third eluate individually to obtain a
15 measure of the amount of said polypeptide in monomeric form relative to the amount of polypeptide in multimeric form in each fraction; and

combining fractions of said third eluate which (almost) exclusively contain the polypeptide in monomeric form to obtain a composition enriched in the polypeptide in the monomeric form.

20 Within the meaning of the invention, the term "a composition that is enriched in the monomeric form of the polypeptide" and the like is any composition, the monomer:multimer ratio of which has been adjusted to conform with the present invention. This might be an untreated cell lysate as obtained following recombinant
25 polypeptide production or a composition which already has undergone some degree of enrichment, but which still does not meet the desired criteria vis a vis the ratio of monomeric to multimeric forms of polypeptide present.

It is contemplated that the "first chromatographic material" and "second
30 chromatographic material" are used as part of a batch process or in a chromatography column. Preferably, chromatography columns will be used. One of ordinary skill in the art will be familiar with the selecting, packing and preparing of such chromatography columns prior to chromatography of proteins.

According to a preferred embodiment of the above method, the first chromatographic material comprising a metal ion is a chromatographic material comprising a divalent metal ion, for example the Ni^{2+} or Zn^{2+} ion. An advantageous first chromatographic material is Fractogel® EMD Chelating (Merck), which has been previously charged with Zn^{2+} . Using such a first chromatographic material, it is advantageously possible to isolate the polypeptide, whether in monomeric or multimeric form, from the extraneous components typically present in, for example, an untreated cell lysate. Co-expression of a functional marker as part of the polypeptide, for example a His-tag or a Flag-tag may facilitate this isolation.

According to another preferred embodiment, the second chromatographic material allows separation on the basis of anion exchange. An advantageous second chromatographic material in this regard is Q Sepharose HP (Amersham Biosciences).

As is typical in protein chromatography, it is advantageous to equilibrate the chromatographic materials, preferably packed into columns, with a buffer prior to actually performing the protein chromatography. After application of the composition or eluate to be isolated or separated to the chromatographic material, this same buffer is used to wash away any material which has failed to bind to the chromatographic material. The volume of first and second buffers used for washing unbound substances from, respectively, the first and second chromatographic materials advantageously corresponds to 6 to 10 times, preferably of 6 times the volume of the respective chromatographic material used. The volume of the running buffer used for translocating substances along the third chromatographic material advantageously corresponds to 1 to 2 times, preferably 1 time the volume of the chromatographic material used. Phosphate buffer (pH 8) is advantageous as both the first buffer and the second buffer, while either phosphate buffer (pH 7.0 - 7.5) or citrate/lysine buffer (pH 6.0 - 7.5) is advantageous as the running buffer.

According to a further embodiment of the inventive method, said method comprises the additional step of analyzing the composition obtained in step (e). In this way, one can obtain a measure of the amount of said polypeptide in monomeric form relative to the amount of polypeptide in multimeric form in the composition. If desired or determined necessary, a further enrichment may ensue by repeating steps (d) through (e). In such a repetition, the composition resulting from the previous round of enrichment is applied to

the third chromatographic material in place of the second eluate. Thus, the process of enriching the monomeric form of the polypeptide such that this form is present in no more than the prescribed or desired proportion within the composition can be an iterative procedure which can be repeated as often as necessary or desired until a given
5 degree of enrichment in the amount of the polypeptide in monomeric form has been reached. Typically, however, one round of enrichment should be sufficient to generate a composition conforming to the criteria set for the composition as defined herein.

It is advantageous to perform such optional analysis using a chromatographic method
10 which separates substances on the basis of their molecular weight . Preferably, such a chromatographic method is high performance size exclusion chromatography performed on an HPLC apparatus. One of ordinary skill in the art will understand how to adjust such HPLC parameters as flow rate, pressure and nature of the mobile phase buffer used. Subsequent analysis by size exclusion HPLC has the advantage that relative
15 amounts of monomeric and multimeric forms of polypeptide can be determined with a high degree of accuracy and sensitivity.

In the inventive method, said imidazole in step (b3) may be applied as a single concentration, or may be applied as a concentration gradient ranging from 60 to e.g.
20 300 mM. Likewise, said sodium chloride in step (c3) may be applied to the second chromatography material as a single concentration, or may be applied as a concentration gradient ranging from 200 to e.g. 500 mM. Such concentration gradients may be a stepwise gradient, i.e. a gradient in which the concentration of, for example, 60 mM imidazole/200 mM sodium chloride is maintained for a period of time before
25 changing to a concentration of, for example, 70 mM/220 mM sodium chloride, which is maintained for a period of time before changing to the next concentration, and so on. The concentration gradient may also be a non-stepwise gradient, i.e. a gradient in which the concentration of imidazole/sodium chloride is increased at a constant linear rate over time. In the event that a single concentration of imidazole is used, advantageous
30 concentrations are 70 mM, 80 mM, 90 mM, 100 mM, 110 mM or 120 mM. In the event that a single concentration of sodium chloride is used, advantageous concentrations are 370 mM, 380 mM, 390 mM, 400 mM, 410 mM or 420 mM.

In an especially advantageous embodiment of the invention, the imidazole is applied in
35 a single concentration of 80 mM to the first chromatographic material. In another

advantageous embodiment of the invention, the sodium chloride is applied in a single concentration of 400 mM to the second chromatographic material. A combination of these advantageous embodiments is particularly preferred. Application of imidazole and sodium chloride in the above respective concentrations has the advantageous effect that the distribution of the monomeric form of the polypeptide and the closest eluting species of the multimeric form of the polypeptide, namely the dimeric form of the polypeptide, are resolved as two distinct, i.e. non-overlapping peaks of polypeptide in the subsequent second separation step (d). Separation of two polypeptide species, here the monomeric and dimeric forms of the polypeptide, with such baseline resolution allows the monomeric form of the polypeptide to be obtained in higher yield free from impurities of the corresponding dimeric form of the polypeptide. This in turn increases the likelihood of obtaining fractions from the second separation containing exclusively or predominantly the polypeptide in monomeric form. As such, the advantageous resolution achieved by the above two concentrations of imidazole and sodium chloride used in concert increases the efficiency with which a composition enriched with respect to the monomeric form of the polypeptide may be obtained.

A further aspect of the invention is a composition (obtainable by the above method of obtaining a composition) which is enriched in the monomeric relative to the multimeric form of the polypeptide. Thus, the method and use of appended claims 20 to 25 may advantageously be carried out/occur with the composition obtained by such above method.

Another aspect of the invention provides a method for the prevention, treatment or amelioration of a proliferative disease, of a minimal residual cancer, of a tumorous disease, of an inflammatory disease, of an immunological disorder, of an autoimmune disease, of an infectious disease, of a viral disease, of an allergic reaction, of a parasitic reaction, of a graft-versus-host disease, of a host-versus-graft disease or of a B cell malignancy. According to this aspect, the composition as disclosed hereinabove is administered to a subject in need of such a prevention, treatment or amelioration.

A further aspect of the invention provides a use of the composition as disclosed hereinabove for the production of a medicament for the prevention, treatment or amelioration of a proliferative disease, of a minimal residual cancer, of a tumorous disease, of an inflammatory disease, of an immunological disorder, of an autoimmune disease, of an

infectious disease, of a viral disease, of an allergic reaction, of a parasitic reaction, of a graft-versus-host disease, of a host-versus-graft disease or of a B cell malignancy.

5 According to a preferred embodiment, the prevention, treatment or amelioration occurs in a human. The tumorous disease is preferably selected from the group consisting of a lymphoma, a B cell lymphoma and a Hodgkin lymphoma. In a further embodiment, the B cell lymphoma is non-Hodgkin lymphoma. In a further embodiment, the autoimmune disease is selected from rheumatoid arthritis, multiple sclerosis, type 1 diabetes mellitus, inflammatory bowel disease, systemic lupus erythematosus, psoriasis, scleroderma and
10 autoimmune thyroid diseases.

Throughout the instant application, it is to be understood that use of a term in the singular may imply, where appropriate, use of the respective term in the plural. Similarly, use of a term in the plural may imply, where appropriate, use of the respective
15 term in the singular.

The invention will now be described further by the appended figures and examples.

Brief description of the figures

- 20 Fig. 1A: Model of a polypeptide comprising two antigen binding sites, wherein one antigen binding site specifically binds the human CD3 antigen, and wherein the polypeptide exists in monomeric form
- 25 Fig. 1B: Model of a polypeptide comprising two antigen binding sites, wherein one antigen binding site specifically binds the human CD3 antigen, and wherein the polypeptide exists in multimeric (here, dimeric) form due to association of two individual target antigen binding sites.
- 30 Fig. 1C: Model of a polypeptide comprising two antigen binding sites, wherein one antigen binding site specifically binds the human CD3 antigen, and wherein the polypeptide exists in multimeric (here, dimeric) form due to association of two individual effector antigen binding sites specific for the human CD3 antigen.

Fig. 2: Up-regulation of the early T cell marker CD69 as a function of concentration of polypeptide in monomeric and multimeric (here, dimeric) form

5 Fig. 3A: Mutual T cell lysis as a function of concentration of polypeptide in monomeric and multimeric (here, dimeric) form using PBMCs as effector cells

10 Fig. 3B: Mutual T cell lysis as a function of concentration of polypeptide in monomeric and multimeric (here, dimeric) form using MC15 cells as effector cells

Examples and detailed description of the figures

15 Example 1: Polypeptide production

Starting from suitable eukaryotic expression vectors, expression of a polypeptide comprising two antigen binding sites is performed in CHO cells in a stirred tank bioreactor using a serum- and protein-free medium. Fermentation is conducted in fed-batch mode at 37°C with glucose feeding. Upon completion of the fermentation process, 20 the supernatant containing secreted polypeptide is harvested by dead end filtration and concentrated 10-fold by cross flow filtration.

The following describes how the ratio of the amount of polypeptide in monomeric form to the amount of polypeptide in multimeric form may be adjusted. As a model for such 25 adjustment, the anti-CD19 x anti-CD3 polypeptide according to SEQ ID NO. 1 (hereinafter "Construct 1") is used, and the multimeric form of Construct 1 is the dimeric form of Construct 1.

Capture of Construct 1 from the cell harvest is performed using an immobilized metal 30 affinity chromatography column (Fractogel EMD Chelating, Merck) charged with zinc (Zn^{2+} -IMAC). The column is equilibrated with 2 column volumes (CV) of phosphate buffer, the cell harvest is applied at 120-180 cm/h and unbound material is washed away with 6 CV of buffer. Applying a step gradient with 60-300 mM Imidazole in phosphate buffer over 5 CV elutes the product. Alternatively, an individual concentration 35 of 70 mM, 80 mM, 90 mM, 100 mM, 110 mM or 120 mM imidazole may be used for this

purpose. Intermediate purification of Construct 1 is performed employing anion exchange chromatography (AEX, Q Sepharose HP, Amersham Biosciences). The column is equilibrated with 2 CV of phosphate buffer pH 8.0 and the eluate from the IMAC column is directly applied to the column. Unbound protein is removed by washing with 6 CV buffer. The product is subsequently eluted with a step gradient of 6 CV of 200-500 mM sodium chloride in buffer. Alternatively, an individual concentration of 370 mM, 380 mM, 390 mM, 400 mM, 410 mM or 420 mM sodium chloride may be used for this purpose. Final adjustment is performed by size exclusion chromatography (SEC) including a separation of monomeric and dimeric forms of Construct 1. A Superdex 200 prep grade column (Amersham Biosciences, bed height >600 mm) is equilibrated with at least 4 CV of either phosphate buffer pH 7.0-7.5 or citrate/lysine buffer pH 6.0-7.5. The sample (corresponding to a volume of 1-5% of the CV) is applied to the column and an isocratic elution using the equilibration buffer is performed. The dimer elutes at approximately 0.5-0.6 CV while the monomer elutes at approximately 0.6 to 0.7 CV (exact elution conditions may vary depending on column length, sample volume, and quality of the column packing). Eluted polypeptide is fractionated and desired fractions are combined. Later fractions contain a higher amount of Construct 1 in monomeric form than do earlier fractions. The ratio of the amount of monomeric Construct 1 to the amount of dimeric Construct 1 may therefore be influenced by the choice of the fraction used.

Specific combinations of elution parameters have proven to be very advantageous. Specifically, elution of the polypeptide, for example Construct 1, from the Zn^{2+} -IMAC column with a single concentration of 80 mM imidazole followed in the next step by elution of this polypeptide from the anion-exchange column with a single concentration of 400 mM sodium chloride yields a mixture of polypeptide which, when resolved by size exclusion chromatography as described above, results in the monomeric form of the polypeptide being baseline-resolved from the next largest multimeric form of the polypeptide, namely the dimeric form of the polypeptide. This lack of overlapping shoulders of peaks corresponding to monomeric and dimeric forms of the polypeptide facilitates the obtaining of fractions containing exclusively or predominantly the monomeric form of the polypeptide; these fractions may later be combined to obtain a mixture in which the content of the monomeric form of the polypeptide has been enriched relative to the content of multimeric or, here, dimeric form of the polypeptide.

As an alternative, cation- or anion- exchange chromatography or chromatography on hydroxyapatite may be used to separate monomeric polypeptide from multimeric, especially from dimeric polypeptide. In both cation and anion exchange chromatography the dimeric form of the polypeptide elutes later during gradient elution. For separation of monomer and dimer using ion exchange, the eluate from the anion exchange column should be diluted. For cation exchange, the pH should be adjusted to allow binding of polypeptide. When using hydroxyapatite chromatography, a low conductivity phosphate buffer should be used.

Analysis of the ratio of relative amounts of monomeric to multimeric polypeptide in a given mixture may be performed by SEC-HPLC using e.g. an Agilent 1100 series HPLC system (or similar). The column used is a Tosoh Biosep TSKgel G3000SWXL column with guard column at a flow rate of 0.6-0.75 mL/minute at a maximum Pressure of 75 bar (7.5×10^6 Pa). As mobile phase a buffer of 100 mM $\text{KH}_2\text{PO}_4/\text{KOH}$, 200 mM Na_2SO_4 pH 6.6 is used. 100 μL of sample are applied. The total run time is 27 minutes. Wavelength of detection is set to 210 nm.

Example 2: Additional biological activities attributable to the polypeptide in multimeric form but not to the polypeptide in monomeric form

A polypeptide comprising two antigen binding sites, one of which specifically binds the human CD3 antigen, is able to bind to (and activate the cytotoxic activity of) cytotoxic T cells via the CD3 antigen located on the surface of such cytotoxic T cells. At the same time, such a polypeptide can specifically bind with its target binding site a surface target on, for example, tumor cells, which would normally not be recognized by cytotoxic T cells. In this manner, the cytotoxic activity of T cells can be directed to, for example, tumor cells as part of a therapeutic regimen to eliminate such cells. Ideally, cytotoxic T cells are only activated upon interaction with a target cell mediated by the polypeptide molecule described above. While the activation mechanism described above seems to be the only biological activity observed for the polypeptide in monomeric form (as defined hereinabove), the polypeptide in multimeric form (as defined hereinabove) has been observed to exhibit additional biological activities.

Polypeptides comprising two antigen binding sites, of which one antigen binding site specifically binds the human CD3 antigen, have a tendency to dimerize.

The following examples therefore discuss the nature of these additional biological activities observed for the polypeptide in multimeric form, using the polypeptide in dimeric form as a concrete example.

Fig. 1A depicts a polypeptide in monomeric form as comprised in the composition of the present invention. The antigen binding sites of the polypeptide are each derived from different antibodies, and each comprises a VH and VL region. The designations "VH/VL" and "VL/VH" denote a mutually exclusive option of either VH or VL at the region so designated. Hence a region designated "VH/VL" would be expected to associate with a region designated "VL/VH" since the two possible associations would result in either, from amino- to carboxy-terminus, VH associating with VL or VL associating with VH. The polypeptide in monomeric form depicted in Fig. 1a would be expected to specifically bind the human CD3 antigen with the left-hand antigen binding site, and another target antigen with the right-hand antigen binding site. The polypeptide may therefore act as a bridge specifically linking a cytotoxic T cell with a target cell of interest while directing the cytotoxic activity of the cytotoxic T cell against the target cell as described hereinabove.

Fig. 1B depicts one possible model for the polypeptide comprised in the present invention, wherein this polypeptide is in multimeric form. Here, the specific polypeptide shown is in dimeric form, meaning that two single polypeptide chains have non-covalently associated to form a homodimeric species. Fig. 1B depicts the scenario in which the two single polypeptide chains have non-covalently associated in an antiparallel fashion through their antigen binding sites which specifically bind target antigen. It should be noted that in this model of dimer formation, the antigen binding sites which specifically bind the human CD3 antigen (each designated "anti-CD3") are free to bind two separate human CD3 antigens (one human CD3 antigen is specifically bound by each anti-CD3 binding site). In contrast, the antigen binding site which specifically binds to target antigen (designated "anti-target") present on one single polypeptide chain is non-covalently associated with the "anti-target" binding site present on the other single polypeptide chain, so that neither of these two target antigen binding sites may specifically bind target antigen. As such, the polypeptide in dimeric form depicted in Fig. 1B would be capable of simultaneously and specifically binding two individual human CD3 antigens, but would be less capable of binding a target antigen.

Fig. 1C depicts another possible model for the polypeptide comprised in the present invention, wherein this polypeptide is in multimeric form. Here, the specific polypeptide shown is in dimeric form, meaning that two single polypeptide chains have non-covalently associated to form a homodimeric species. Fig. 1B depicts the scenario in which the two single polypeptide chains have non-covalently associated in an antiparallel fashion through their effector binding sites which specifically bind the human CD3 antigen. It should be noted that in this model of dimer formation, the antigen binding sites which specifically bind the target antigen (each designated "anti-target") are free to bind two separate target antigens (one target antigen is specifically bound by each anti-target binding site). In contrast, the antigen binding site which specifically binds to the human CD3 antigen (designated "anti-CD3") present on one single polypeptide chain is non-covalently associated with the "anti-CD3" binding site present on the other single polypeptide chain, so that neither of these two antigen binding sites may specifically bind the human CD3 antigen. As such, the polypeptide in dimeric form depicted in Fig. 1C would be capable of simultaneously and specifically binding two individual target antigens, but would be less capable of binding a human CD3 antigen.

Example 2a: Activation of T cells by polypeptide in multimeric (here, dimeric) form in the absence of target cells

Peripheral blood mononuclear cells (PBMCs) were prepared from blood of a healthy donor by Ficoll density centrifugation. To investigate whether the polypeptide of the inventive composition in multimeric (here, dimeric) form is capable of activating T cells in the absence of target cells, PBMCs were incubated with a polypeptide comprising two antigen binding sites. One antigen binding site (the effector binding site) of the polypeptide specifically bound the human CD3 antigen, and the other antigen binding site (the target antigen binding site) of the polypeptide specifically bound the human EpCAM antigen. This particular polypeptide was chosen for study because the interaction with target cells could be excluded due to the absence of EpCAM-positive cells in the PBMC population; any effects observed in using the above polypeptide with PBMCs would be attributable solely to the binding site specifically binding the human CD3 antigen.

In order to compare the effect of polypeptide in monomeric form with the effect of polypeptide in dimeric form, the polypeptide had previously been resolved into fractions containing either exclusively monomeric polypeptide (as for example modeled in Fig.

1A) or exclusively dimeric polypeptide (as for example modeled in Figs. 1B and 1C). Resolution of polypeptide into these fractions was accomplished as described above in Example 1.

5 In round-well microtiter plates, 2×10^5 PBMCs/well were incubated in a volume of 200 μ L with either pure monomer or pure dimer fractions of the polypeptide at the concentrations indicated in Fig. 2. Using flow cytometry, the expression levels of CD69 were analyzed in each sample after a 24-hour incubation period. CD69 is a marker on the surface of T cells, the up-regulation of which can serve as an early indicator of T cell
10 activation. By monitoring the expression levels of CD69 in the various samples, it is possible to obtain an early measure of the degree to which the activation of T cells has taken place. T cells were identified with an anti-CD3-specific antibody. Samples were analyzed in duplicate. As can be seen in Fig. 2, incubation with the polypeptide in dimeric form resulted in more than 20 % of T cells being activated at a polypeptide
15 concentration of 1 μ g/mL. The lowest concentration of polypeptide in dimeric form eliciting an expansion of CD69-positive T cells was 10 ng/mL. In contrast, the polypeptide in monomeric form induced CD69 expression of only about 3 % of the T cells at the highest tested concentration (1 μ g/mL of polypeptide in monomeric form). The minimal degree of activation observed in response to the polypeptide in monomeric
20 form at a concentration of 1 μ g/mL might be a result of residual polypeptide in dimeric form still present in the preparation of polypeptide in monomeric form. These data demonstrate that the polypeptide in dimeric form is able to activate T cells in the absence of target cells while the monomer is not. This capability represents an activity other than the killing of target cells which is attributable to the polypeptide in dimeric
25 form but not to the polypeptide in monomeric form.

Example 2b: Mutual T cell lysis by polypeptide in multimeric (here, dimeric) form

To analyze whether the polypeptide in multimeric (here, dimeric) form is capable of killing T cells two sets of experiments were performed in which effector cells were co-
30 incubated with the T cell line HPBALL (DSMZ No ACC 483; DSMZ = Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH) in the presence of polypeptide. In the first set of experiment, PBMCs were used as the effector cells, whereas the effector cells used in the second set of experiments were MC15 cells (Biesinger B., Müller-Fleckenstein I., Stimmer B., Lang G., Wittmann S., Plater E.
35 Desrosiers R.C. and Fleckenstein B.; 2002, Proc. Natl. Acad. Sci. USA, 89, 3116-3119).

The polypeptide used for this experiment comprised two antigen binding sites. One antigen binding site (the effector binding site) specifically bound the human CD3 antigen, and the other antigen binding site specifically bound the human CD19 antigen, a pan-B cell marker described hereinabove. HPBALL cells have been described to be CD3-positive. Blood cells were washed out from Leukocyte filters. PBMCs were prepared by Ficoll density centrifugation. MC15 cells were cultured as described in the literature reference above in this paragraph. To distinguish the effector cells from target cells, the HPBALL cells were stained with the fluorescent dye Calcein AM according to the manufacturer's protocol.

In order to compare the effect of polypeptide in monomeric form with the effect of polypeptide in dimeric form, the polypeptide had previously been resolved into fractions containing either exclusively monomeric polypeptide (as for example modelled in Fig. 1A) or exclusively dimeric polypeptide (as for example modelled in Figs. 1B and 1C). Resolution of polypeptide into these fractions was accomplished as described above in Example 1.

In round-bottom microtiter plates, 5×10^5 effector cells were incubated with 5×10^4 HPBALL cells for 4 hours in the presence of either highly pure monomer or highly pure dimer fractions of the above polypeptide at the concentrations indicated in Fig. 3A (for PBMC effector cells) and Fig. 3B (for MC15 effector cells). Appropriate controls containing HPBALL cells and effector cells were incubated in the absence of polypeptide. After the incubation period the supernatants were harvested. The amount of fluorescent dye released by dead cells was measured using a Spectrafluorometer. As can be seen in each of Fig. 3A and Fig. 3B, the polypeptide in dimeric form induced HPBALL cell lysis at concentrations higher than 10 ng/mL. In contrast, no target cell lysis by the polypeptide in monomeric form was observed under identical conditions. This finding demonstrates that lysis of CD3-positive cells has occurred and is attributable to the polypeptide in dimeric form but not to the polypeptide in monomeric form.

Example 3: General propensity of polypeptides to form dimers

It was desired to show that the propensity to form a multimeric species is common to the general class of bispecific single chain antibodies in which one binding specificity is for the human CD3 antigen. To this end, a number of such bispecific antibodies were produced in Chinese hamster ovary (CHO) cells according to generally known

procedures (Sambrook et al., 1989). Each bispecific single chain antibody produced contained two antigen binding sites, each antigen binding site containing one VH and one VL antibody region. One of the two antigen binding sites in each molecule was specific for the human CD3 antigen. The other antigen binding site ("target antigen binding site") was specific for a desired target antigen other than the human CD3 antigen. Proportions of polypeptide in monomeric and multimeric (here, dimeric) form were determined by a combination of SDS-PAGE performed under reducing conditions, Western Blot performed using Penta-His (Qiagen) and Goat-anti-mouse-AP (Sigma) antibodies and gel filtration performed on a Sephadex S200 column. The relative proportions of bispecific single chain polypeptide present in dimeric form are shown below in Table 1 for polypeptides comprising target antigen specificities against the human CD19 antigen, the human EpCAM antigen, the human Wue1 antigen (a highly specific multiple myeloma antigen) and the human sTn antigen (a carbohydrate displayed on the epithelium of malignant cells in breast, prostate and colon cancers).

Table 1

Target antigen specificity	Approximate % of single polypeptide chains present as a monomer	Approximate % of single polypeptide chains present as a dimer
CD19 ("Construct 1" from above)	~ 65-70 %	~ 30-35 %
EpCAM	~ 75 %	~ 25 %
Wue1	~ 85-90 %	~ 10-15 %
sTn	~ 75-80 %	~ 20-25 %

As can clearly be seen in Table 1, each bispecific single chain antibody with anti-human CD3 antigen binding specificity spontaneously forms significant amounts of multimeric (i.e. here, dimeric) species when left uncontrolled. The propensity to spontaneously form homodimers therefore appears to be a generic characteristic of the class to which the bispecific single chain antibodies examined here belong.